

# A mouse brain cDNA encodes a novel protein with the protein kinase C phosphorylation site domain common to MARCKS

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We have isolated a mouse brain cDNA clone encoding a protein of 200 amino acids ( $M_r$  20165) with partial homology with MARCKS (myristoylated alanine-rich C-kinase substrate). Two regions show similarity with MARCKS, one is the kinase C phosphorylation site domain which is supposed to bind calmodulin, and the other is the region near to the N-terminus, including the consensus sequence of myristoylation. It has a similar amino acid composition to MARCKS, but the content of alanine is not as high. It is distributed throughout the mouse brain, but the pattern is not identical with that of MARCKS. Both proteins may be members of a new protein family involved in coupling the protein kinase C and calmodulin signal transduction systems.

Myristoylated alanine-rich C-kinase substrate; Protein kinase C; Calmodulin

## 1. INTRODUCTION

Protein kinase C is a key enzyme of intracellular signal transduction [1]. Although the enzyme itself is well-characterized, little is known about its physiologically important substrates. One of the few examples is myristoylated alanine-rich C kinase substrate (MARCKS) [2], formerly known as 87-kDa protein [3–6]. This protein is found in a wide variety of cell types, and is phosphorylated within seconds by protein kinase C activation, in the same manner as growth factors in fibroblasts, neurotransmitters in neuronal and glial cells, as well as phorbol-esters or cell-permeating diacylglycerols. In macrophages, part of the protein is myristoylated and found associated with membranes, while the phosphorylated form is found mainly in cytosol [7]. In isolated nerve terminals, phosphorylation by kinase C leads to the translocation of MARCKS from the membrane to the cytosol [8]. It binds calmodulin, and this binding can be prevented by protein kinase C-catalyzed phosphorylation of the protein [9].

cDNA clones encoding MARCKS have been recently isolated, and sequenced [2,10]. The deduced proteins have acidic isoelectric points, and an unusual high content of alanine and glutamic acid. The sites phosphorylated by kinase C have also been determined [11]. All of these are in a 25 amino acid domain, which is extremely basic. A synthetic peptide corresponding to

this domain binds calmodulin with high affinity, and phosphorylation of this peptide inhibits the binding [9]. This result strongly suggests that this domain is responsible for the main properties of MARCKS.

In this report, we describe a mouse brain cDNA which encodes a protein with the kinase C phosphorylation site domain of MARCKS. Near the N-terminus there is another region which can be identified, because it includes the consensus sequence of myristoylation. The mRNA is distributed throughout the mouse brain, but the pattern is not identical with that of the MARCKS protein [12]. Thus these two proteins may be members of a new protein family involved in coupling the protein kinase C and calmodulin signal transduction systems.

## 2. MATERIALS AND METHODS

The isolation of the partial cDNA clone of F52 has previously been described [13]. Total RNA was extracted by the guanidinium-CsCl method from mouse brain, and poly(A)<sup>+</sup> RNA was purified by oligo(dT) cellulose column chromatography [14]. A  $\lambda$ gt10 mouse brain cDNA library was constructed using the method of Gubler and Hoffman [15]. To obtain full-length copies, the cDNA was fractionated on a Sepharose CL-4B column, and the top 5% was used for the library. Among several positives, the longest clone was selected and sequenced by the dideoxy chain termination method [16] using a shot-gun strategy with oligowalking.

Homology searches used the FASTA program [17]. The amino acid alignment of the homologous amino acid sequence was done using Clustal [18].

Mouse brain sections were mounted on organosilane-treated microscope slides and stored at  $-20^{\circ}\text{C}$  until use. Following fixation and pronase digestion, the sections were prehybridized for 2 h at  $37^{\circ}\text{C}$  in 50% formamide,  $5 \times \text{SSC}$ , 10 mM sodium phosphate (pH 7.0),  $2 \times \text{Denhardt's}$ , 0.1% SDS. Hybridization was performed at  $42^{\circ}\text{C}$  for 30 to 40 h in the above solution containing 250  $\mu\text{g/ml}$  denatured salmon sperm DNA and  $^{32}\text{P}$ -labelled DNA probe ( $1\text{--}1.5 \times 10^6$  cpm/ml). The probe used was single stranded DNA derived from

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1	GGAGCGCAGCAGCAGTGGGCGCAGCTAGCGGGTTCGGCTGCGGAGCGGGGGTGCAGCGCGC	60
61	TTTGGCCCCCGTACCCCTCCCCCTCGGACGCAGCCCCACCCCTCTGCCCGCCGGGCGCGTC	120
121	CCAGCCGAACCTATCCCCTGCGGCGCCAGCGCGGGGCGGCTCCGGGCGCCCCCAGCAGACC	180
-1	M G S Q S S K A P R G D V T A E E A	18
181	CCCATCATGGGCAGCCAGAGCTCTAAGGCTCCCCGGGGCGACGTGACCGCCGAGGAGGCA	240
19	A G A S P A K A N G Q E N G H V R S N G	38
241	GCGGGCGCTTCCCCCGCGAAGGCCAACGGACAGGAGAATGGCCACGTGAGAAGCAATGGA	300
39	D L T P K G E G E S P P V N G T D E A A	58
301	GACTTAACCCCAAGGGTGAAGGGGAGTCACCACCCGTGAACGGAACAGATGAGGCAGCT	360
59	G A T G D A I E P A P P S Q E A E A K G	78
361	GGGGCTACTGGTGTATGCCATCGAGCCAGCGCCCCCTAGCCAGGAAGCTGAGGCCAAGGGG	420
79	E V A P K E T P K K K K K F S F K K P F	98
421	GAGGTCGCCCCCAAGGAGACCCCCAAGAAGAAGAAATTCTCTTTCAAGAAGCCTTTC	480
99	K L S G L S F K R N R K E G G G D S S A	118
481	AAATTGAGTGGCCTGTCCTTCAAGAGAAATCGGAAGGAGGGTGGGGGTGATTCTTCAGCC	540
119	S S P T E E E Q E Q G E M S A C S D E G	138
541	TCCTCGCCAACAGAGGAAGAGCAGGAGCAGGGCGAGATGAGCGCCTGCAGCGACGAGGGC	600
139	T A Q E G K A A A T P E S Q E P Q A K G	158
601	ACTGCCCAAGAAGGGAAGGCCGCTGCCACCCCTGAGAGCCAAGAGCCCCAGGCCAAGGGG	660
159	A E A S A A S K E G D T E E E A G P Q A	178
661	GCAGAGGCTAGTGCTGCCTCCAAGGAAGGAGACACAGAAGAGGAGGCAGGGCCCCAGGCT	720
179	A E P S T P S G P E S G P T P A S A E Q	198
721	GCAGAGCCATCCACTCCCTCGGGCCAGAGAGTGGCCCCACACCTGCCAGTGCTGAGCAG	780
199	N E *	218
781	AATGAGTAGGGGTGGGGACAGGTGGGTGATCTCTAAGCTGCAAAAAGTGTGCTGTCTTG	840
841	TGAGGTCAGTGCCTGGACCTTGTGCCCTGGCTGCCTTCTTGTGCCAGAAAGGAAGGGGC	900
901	TTGTTGCCCCCTCCAGCCACGCTCCCTTTTTTGCCCTCCTGTGGATTCTCCCATCAGCCA	960
961	TTTGGTCTTACTCTTAAGGCCAGTTGAAGATGGTCCCTTACGGTTTCCCAAGTTAGGTTA	1020
1021	GTGATGTGAGATGCCCTGTCCCTACCTCCTTCCCGAGCCCCATCCCTGCAGAAGGCAATT	1080
1081	GCTGGGTTTCCAATTCTTTTCCAAGTTCGTTTTGTTTTATCCTGCTTCTCAACCCCTGAG	1140
1141	CCAGAAGTGGGGGCTTATACTCCCAAACCTGAGTGTCCAGCCTTTCCCTGTTGAATTC	1200
1201	TTAGTCTCTTGTGCTGTGCCTAGTGGCACCTGGGCTGGGGAGGACATTGCCCATCTGGG	1260
1261	TTTTTATAAATGTCTTACTCAAGTTCAAACCTCCAGCTTGTGAAATCAACCATGTCCCTG	1320
1321	ACTTGATAAGCAAGTGTAGGCTTCAGGAGGGAGGGAGGTCTGTAATGTGAAACAACCTC	1380
1381	TTGTTGTCTTTTTTTTTTCCCCCTCCCATTTGTTGTAATAACTTTTAATGGCCAAACCCAG	1440
1441	ATTTGTACTTTTTTTTTTTTTTCTAACTGCTAAAACCATTTCTCCACCCGGATTTACTGTA	1500
1501	ACATTTGAAAAGGAATAAATGTCGTCCCTTTAAAAA	1560
1561	AAAAAAAAAAAAAAAAAAAAA	1584

Fig. 1. Nucleotide and predicted amino acid sequence of a cDNA encoding the F52 protein, a protein with the phosphorylation site domain of MARCKS. The amino acid sequence of the predicted polypeptide is shown as single letter code above the nucleotide sequence.

a M13 clone containing the F52 insert. The sections were washed in three changes of  $2 \times$  SSC at room temperature, and then in 50% formamide, 0.5 M NaCl, 10 mM Tris-HCl, 1 mM EDTA (pH 7.4) at 37°C for about 6 h. They were dehydrated in 70% ethanol, and exposed on a Kodak X-AR 5 film.

### 3. RESULTS

In a previous experiment, 35 cDNA clones with specific expression patterns in mouse brain were isolated from 950 cDNA clones by Northern and in situ hybridization using the cDNA inserts as probes [13]. DNA sequence analysis of full-length clones revealed that one, F52, had an open reading frame encoding 200 amino acids (Fig. 1). A comparison of the predicted amino acid sequence with the existing data base revealed that F52 was related to myristoylated alanine-

rich C kinase substrate (MARCKS). The amino acid alignment of F52, bovine MARCKS [2] and chicken MARCKS [10] is presented in Fig. 2. Similarity is observed in only two regions, from the first methionine to position 47, and from position 152 to 176 (Fig. 2). The first region includes the consensus sequence of myristoylation [19,20] around glycine 2, which may be myristoylated. The second region corresponds to the 25 amino acid domain with phosphorylation sites for protein kinase C, with conservation of three of the four serines. The domain maintains a strong basic nature, in spite of some amino acid substitutions. The overall amino acid identity is as follows: F52 and bovine MARCKS, 52%; F52 and chicken MARCKS, 52%; bovine and chicken MARCKS, 65%.

		20	40	60
F52	MGSQSSK-APRGDVTAEAA-----GASPAKANGQENGHVRSNGDLTPKGE-----			
MARCKS (BOVINE)	MGAQFSKTAAKGEATAERPGEAAVASSPSKANGQENGHVKVNGDASPAAAEPGAKEELQA			
MARCKS (CHICKEN)	MGAQFSKTAAKGEAAAEKPGEA-VAASPSKANGQENGHVKVNGDASPAAAEAG-KEEVQA			
		** * * * *	** * * * * *	** * *
		80	100	120
F52	-----GESPPVNGTDEAAGAT-----GDAIEPAPPSQ-			
MARCKS (BOVINE)	NGSAPAADKEEPAAAGSGAASPAAAEKDEPAAAPDAGHPVEKEAPVEGGAAEPGSPTAA			
MARCKS (CHICKEN)	NGSAPAE-----TGKEEAASSEPAS-----EKEAA-EAESTEPASP--A			
			* *	** *
		140	160	180
F52	EAEAKGEVAPKETP-----KKKKKFSFKKPFKLSGLSFKRNRKEGG--			
MARCKS (BOVINE)	EGEAASAASSSSSPKAEDGGHALAQQRDAEKKKKRFSFKKSFKLSGFSFKKNKKEAGEG			
MARCKS (CHICKEN)	EGEA-----SPKTEEGATP-SSSETPKKKKKRFSFKKSFKLSGFSFKKNKKEAGEG			
		* **	**** * * * *	* * * *
		200	220	240
F52	-----GDSSASSPTEEEQEQGEMSACSDEGTAQEGKAAATPESQEPQAKGAEASAASKEGD			
MARCKS (BOVINE)	GAEAGAAGASAEGGKDEASGGAAAAAGEAGAAPGEPTAAPGEEAAAGEEGAAGGDPQEA			
MARCKS (CHICKEN)	AESEGGAAAAAEGGKEEAAAAPEAAGGEEGKAAAEASAAAAGSREAAKEEAGDSQEA			
		* *	*	*
		260	280	300
F52	TEEEA-----GPQAAE-----			
MARCKS (BOVINE)	PEEAAVAPEKPPARRGAKAVEEPSKAEKAEAEAGVSAAGAAGCEAPSAAGPGCPRAGGAP			
MARCKS (CHICKEN)	SDEAA--PEKATGE-----EAPAAEQQQQQQEQKAAEEAGAAATSEAGSG--EQEAAP			
		* *	*	
		320		
F52	-----PSTPSGPESGPTPASAEQNE			
MARCKS (BOVINE)	REEAAPPRASSACSAPSQEAQPECSPEAPPAAEAE			
MARCKS (CHICKEN)	AEEPAAAR-----QEAPSESSPEGP-AEPAE			
			* *	** *

Fig. 2. Comparison of the amino acid sequence of F52, bovine and chicken MARCKS. Identical residues are indicated by asterisks. Putative phosphorylation sites are indicated by arrowheads. Note that a high degree of conservation is only observed in the phosphorylation site domain and the region near the N-terminus.

The predicted molecular weight of F52 is 20165, which is smaller than the predicted molecular weights of 31979 and 27728, for bovine and chicken MARCKS, respectively. F52 has a high proportion of alanine (15.5 mol%), but this is not as high as the 28.4 and 27.0% found in bovine and chicken MARCKS. F52 has other features of the amino acid composition of MARCKS. All proteins show a high amount of glutamic acid and glycine.

An in situ hybridization experiment was performed to determine the expression sites of F52 in mouse brain. Although F52 is expressed throughout the mouse brain, it is enriched in the following regions: dentate gyrus, anterior olfactory nucleus, primary olfactory cortex, entorhinal cortex, medial preoptic area, and dorsomedial hypothalamic nucleus (Fig. 3).

#### 4. DISCUSSION

One approach to understand the consequences of the activation of protein kinase C is to characterize cellular substrates of protein kinase C, of which MARCKS is the best example. Phosphorylation by protein kinase C regulates its binding capacity to calmodulin; it has a myristoylation site, explaining its localization in the membrane of macrophages or nerve terminals. The domain with phosphorylation sites is highly basic and has a general  $\alpha$ -helical conformation, similar in certain respects to the calmodulin-binding regions of other proteins [21].

F52 resembles chicken and bovine MARCKS with strong conservation of a phosphorylation site domain. It is likely that this is functionally important, and F52

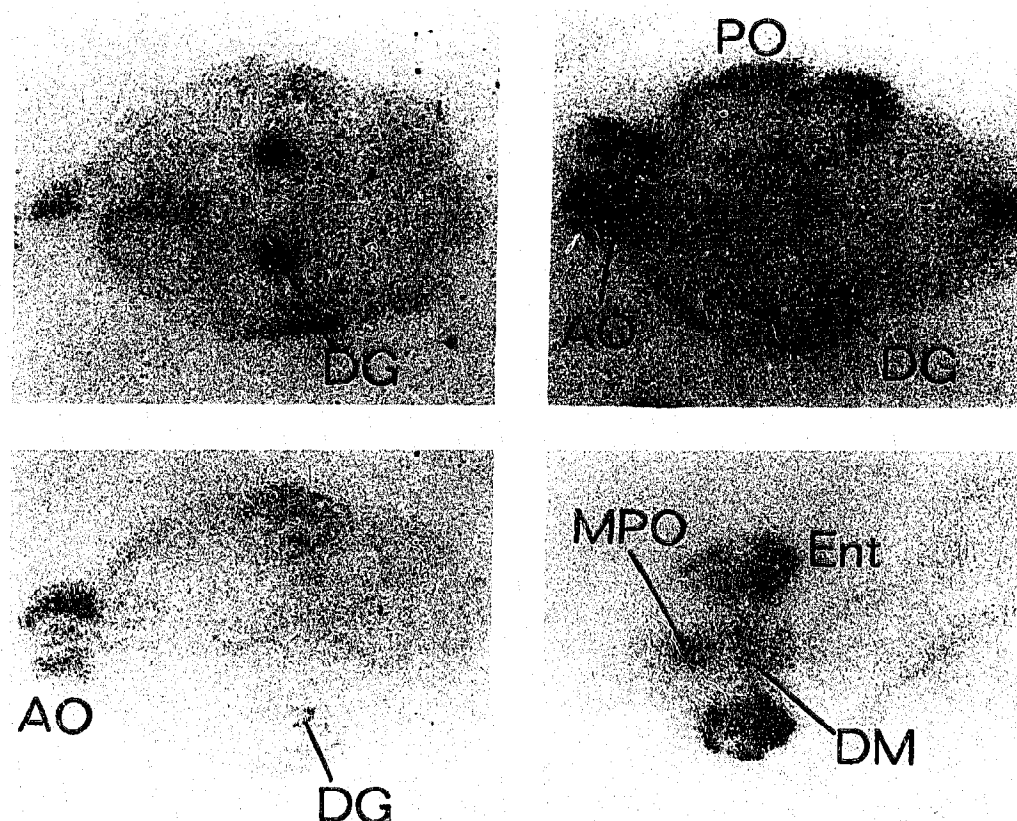


Fig. 3. In situ hybridization analysis of F52 in the mouse brain. The probe used is the partial cDNA clone of F52 which was also used for the isolation of the full-length copy. Abbreviations are as follows: AO, anterior olfactory nucleus; Ent, entorhinal cortex; DG, dentate gyrus; DM, dorsomedial hypothalamic nucleus; MPO, medial preoptic area; PO, primary olfactory cortex.

is also a calmodulin-regulating protein. The divergence of the rest of the structure from that of MARCKS suggests that the rest is probably not relevant to this function. This is supported by existence of another homologous region at its N-terminus, which includes the consensus sequence of myristoylation. This suggests that F52 also could be localized to the membrane.

MARCKS and F52 are distributed throughout the brain, but the enriched regions are not identical [12]. Both are enriched in primary olfactory cortex, entorhinal cortex and hypothalamus. While MARCKS is enriched in amygdaloid complex, F52 is enriched in anterior olfactory nucleus and dentate gyrus. Although it may be misleading to compare the data of different techniques (MARCKS, immunocytochemistry; F52, in situ hybridization) and of different species (MARCKS, rat; F52, mouse), the cellular subpopulations which express F52 seem different from those which express MARCKS.

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